

Marcus Zervos, MD

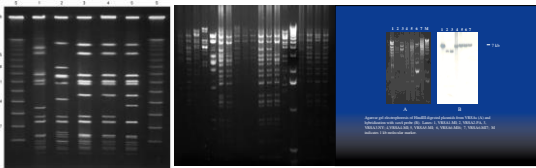
Are Clinical Labs Ready for Emerging Infectious Diseases

Professor of Medicine and Assistant Dean of Global Affairs,
Wayne State University School of Medicine
Division Head, Infectious Diseases, Henry Ford Health

The Big Picture

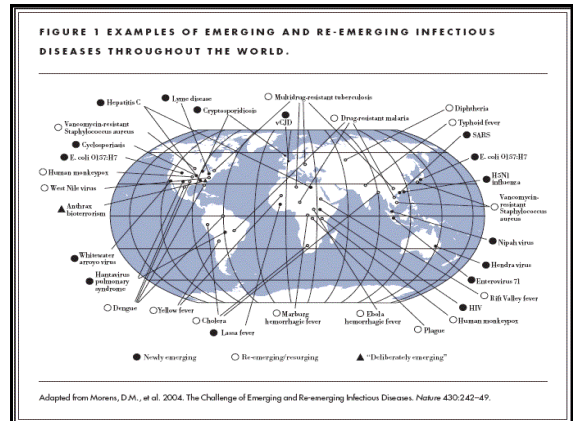
- Transformational advancements in field of diagnostics
- Where we are/where have we been
- Where are we going

Where have we been/where are we



Zervos M 1987 Ann Int Med
<https://doi.org/10.7326/0003-4819-106-5-687>

Smith T NEJM 1999; 340:493-501
DOI: 10.1056/NEJM199902183400701



57,527 cases global, 21,894 USA

Monkeypox



<https://www.cdc.gov/poxvirus/monkeypox/index.html> accessed 9.9.2022

Poliovirus

- June 2022, poliovirus was confirmed in an unvaccinated immunocompetent adult resident of New York hospitalized with flaccid lower limb weakness.
- Vaccine-derived poliovirus type 2 was isolated from the patient and identified from wastewater samples in two neighboring New York counties.

Link-Gelles et al MMWR August 19,2022;71(33):1065-8

Where are we going

- Outbreak investigation
- Public health surveillance
- Clinical applications



Genomics can provide powerful data in real time

- Informing diagnostics, vaccines and therapies
- Identifying variation across the genome
- Tracking and characterizing outbreak transmission
- Getting to functional mutations
- Informs countermeasures (prevention and therapy)



Molecular/new tests

- Detect:
 - what organisms are out there/tests for emerging diseases/known and novel pathogens
 - test for multiple agents
 - rapid
- Connect: information/data
- Empower: everyone in system/partner/training
 - The community to improve preparedness, response

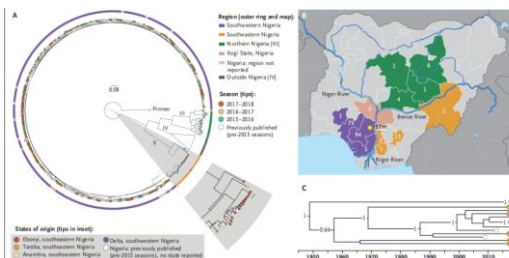


Partnership

- Clinicians need to partner with laboratory:
 - What tests are implemented/available
 - Who to test
 - How to interpret/report comments
 - Who can order
 - Develop with lab algorithms and guidelines for use, restrictions on ordering



Outbreak investigation Lassa fever: Nigeria



Siddle KJ NEJM 2018;379:1745-53



Real-Time Genomic Surveillance

- 413 isolates at University of Pittsburgh
 - Detected 18 *P. aeruginosa*, pseudo outbreak of *Serratia* and a cluster of VRE
- CEID lab detected
 - cluster of NDM in a nursing home
 - in 89 *S. aureus* cluster of daptomycin resistant *S. aureus*
 - SARS-Co-V2 651 samples

Sundermann A Clin Infect Dis 2021;doi:10.1093/cid/ciab946

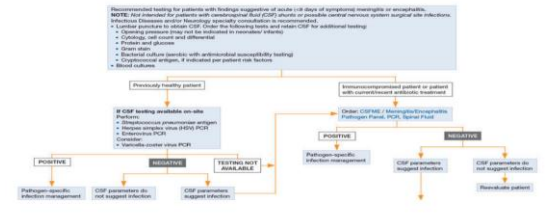


Molecular methods to detect and differentiate MSSA and MRSA .

- Is becoming a useful tool for rapid detection of bacteremia caused by MSSA and MRSA.
- Requires an active stewardship team
 - The median time to appropriate therapy was significantly shorter in the post-RDT group (15 versus 0 h, $P < 0.001$), and the mean length of stay for patients with coagulase-negative staphylococcus was significantly shorter (10.5 versus 7.7 days; $P = 0.015$).
- Useful for infection control

Bukowski PM 2018 62(11) e01334-18

Meningitis/Encephalitis Panel Algorithm



The Big Picture

- Transformational advancements in field of diagnostics
 - WGS
 - Sanger sequencing
 - Metagenomics, next gen sequencing
 - CRISPR
 - Biomarkers
 - Multiplex biosensing technologies

WGS

- Allows opportunity to study the evolution in real time
- How selective pressures vary over time and affect vaccine and treatment, transmission virulence, immunity especially in immune compromised
- Use for surveillance and outbreaks/impacts public health

COVID-19 highlights the need for testing and genomic capacity

- Establish testing sites
- Scaling up sequencing capacity
- Develop cloud based analytics
- Investigating transmission, variants associated with transmission, vaccine and treatment failure

WGS application to SARS-CoV-2

- Study of the ability of the virus to evolve, the traits that have been selected and the genomic regions responsible for them.
- How selective pressures vary over time, as immunity from vaccines or prior infection accumulates in the population
- The importance of monitoring long term infections in immunocompromised patients.

COVID-19 What have we learned

- We were not prepared: erosion of public health infrastructure
- Public health cannot just respond to emergencies
- Health inequity
- Build laboratory capacity
- Standardize and improve data systems/analytics/big data
- Translate research into programs/learn from response
- Improve communication/utilize media
- Build public health staffing/workforce/collaborations
- Living guidance for clinical management



We were not prepared: early testing



Sanchez et al MMWR 2020;69:27:882-6



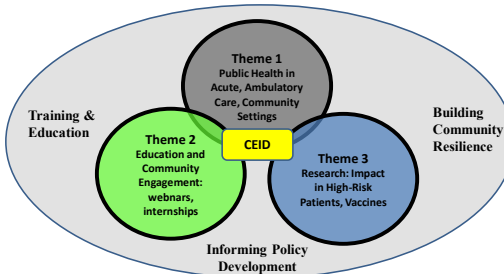
Transitioning to Endemic State: The Pandemic is not over

- Many similarities to pandemic influenza. Over time, influenza became a manageable illness with vaccines and effective therapies
- We can optimize our public health response by:
 - Building up laboratory surveillance
 - Enhancing ability to provide rapid testing
 - Conducting prompt contact tracing and outbreak investigation and control
 - Addressing vaccine hesitancy and delivery issues



Genomic Surveillance Laboratory: MI-SAPPHIRE

- CEID received \$4.3 million in federal funding to expand sequencing for COVID-19 and other infectious diseases
- CEID will collect and analyze genomic data to address emerging infectious disease threats and enhance the city, state and region's ability to respond to those threats
- CEID will increase lab and sequencing capacity in Detroit starting with SARS-CoV-2 and then other infectious disease threats with the potential for broad community spread.
- Funding for the Michigan Sequencing Academic Partnership for Public Health Innovation and Response (MI-SAPPHIRE) is through a CDC Epidemiology and Laboratory Capacity grant MDHHS received.



Platforms: Building public health and research laboratory capacity, testing, vulnerable populations, bioinformatics, clinical trials, community-based participatory research, electronic health records, genomics, mobile health, practice-based research networks, surveillance, outbreak prevention



Metagenomics

- Metagenomic next-generation sequencing (mNGS) is a shotgun sequencing approach in which all of the nucleic acid (DNA and RNA) in a clinical sample is sequenced at a very high depth, 10-20 million sequences per sample.
- mNGS can be performed for any type of clinical sample, including cerebrospinal fluid, plasma, respiratory secretions, saliva, urine, stool, or tissue

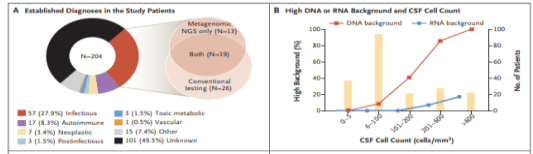


Metagenomics

- Can be used for novel pathogen discovery/public health surveillance
- Clinical use CLIA validated
- Amenable to POC
- Relatively rapid
- ID by host response to particular condition or pathogen (eg West Nile not detected by PCR, febrile illness infection vs autoimmune)



Clinical Application Metagenomics



Wilson MR et al
NEJM
2019;380:2327-40

Metagenomics:UCSF Lab

Representative Set of mNGS Assay Positives (2016-2022)

Streptococcus agalactiae (CSF)	Angiostromyxa cantonensis (CSF)	Powassan virus (CSF)
Enterobacter aerogenes (CSF)	rubella virus* (CSF)	human bocavirus 1 (plasma)
Human herpesvirus 1-8 (CSF, plasma)	yellow fever virus* (CSF)	Legionella anisa (plasma)
Cryptococcus neoformans (CSF)	Petavi virus* (CSF)	Candida parapsilosis (plasma)
enterovirus D68 (CSF)	astrovirus*** (CSF)	Gardnerella vaginalis (plasma)
JC polyomavirus (CSF)	Colorado tick fever virus (CSF)	various bacterial species in sepsis (plasma)
SARS-CoV-2 (respiratory fluid)*	Babesia microti (plasma)	Coccidia burnetti (plasma)
Candida sp. (CSF)	Naegleria fowleri (CSF)	Fusarium sp (plasma)
Anaplasma sp. (CSF)	Acinetobacter sp. (CSF, plasma)	parvovirus B19 (plasma)
Sarbecovirus sp. (CSF)	Chlamydia psittaci (CSF)	Legionella pneumophila (CSF)
St. Louis encephalitis virus (CSF)	Brucella melitensis (CSF)	Mycobacterium sp. (CSF, plasma, respiratory fluid)
hepatitis E virus (CSF)	Mycoplasma pneumoniae (plasma)	Klebsiella kingae (plasma)
Pasteurella marshallii (CSF)	respiratory viruses (respiratory fluid)*	BK polyomavirus (plasma)
HIV polyomavirus (CSF)	Cache Valley bunyavirus (CSF)	Bartonella quintana (plasma)
Capnocytophaga canimorsus (plasma)	tularemia (plasma)	

[For Providers \(ucsf.edu\)](#)

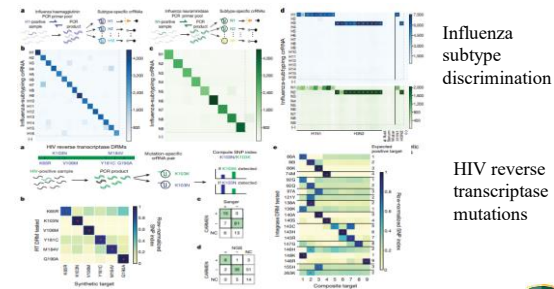


CRISPR

- Repeat sequences/spacers transcribed into RNA or DNA, which are complimentary to viral genome
 - CRISPR allows bacteria to defend themselves vs phages
- Cas13 for RNA viral diagnostics
- Amenable to microassay, multiplex, POC, relatively rapid
- Doesn't need informatics



CRISPR detection using cas13



Conclusion

- We have the power to prevent and defeat infectious diseases, but only we step up the fight
- We must pool our greatest resources—our imagination and intellect—to fight this collective fight. For as Joshua Lederberg noted, "Pitted against microbial genes, we have mainly our wits."
- We need a vision a strategy and a plan for diagnostics that can meet the challenges of the 21st century

